Pedosphere **30**(1): 18–24, 2020 doi:10.1016/S1002-0160(19)60826-X ISSN 1002-0160/CN 32-1315/P \circ 2020 Soil Science Society of China Published by Elsevier B.V. and Science Press

PEDOSPHERE

www.elsevier.com/locate/pedosphere

A new perspective on functional redundancy and phylogenetic niche conservatism in soil microbial communities

Yu JIA and Joann K. WHALEN*[∗]*

Department of Natural Resource Sciences, Macdonald Campus of McGill University, Ste-Anne-de-Bellevue, Quebec H9X 3V9 (Canada)

(Received January 4, 2019; revised June 17, 2019)

ABSTRACT

Functional redundancy in soil microbial communities seems to contradict the notion that individual species have distinct metabolic niches in multi-species communities. All soil microbiota have the metabolic capacity for "basic" functions (*e.g.*, respiration and nitrogen and phosphorus cycling), but only a few soil microbiota participate in "rare" functions (*e.g.*, methanogenesis and mineralization of recalcitrant organic pollutants). The objective of this perspective paper is to use the phylogenetic niche conservatism theory as an explanation for the functional redundancy of soil microbiota. Phylogenetic niche conservatism is defined as the tendency for lineages to retain ancestral functional characteristics through evolutionary time-scales. The present-day soil microbiota is the result of a community assembly process that started when prokaryotes first appeared on Earth. For billions of years, microbiota have retained a highly conserved set of core genes that control the essential redox and biogeochemical reactions for life on Earth. These genes are passed from microbe to microbe, which contributes to functional redundancy in soil microbiota at the planetary scale. The assembly of microbial communities during soil formation is consistent with phylogenetic niche conservatism. Within a specific soil, the heterogeneous matrix provides an infinite number of sets of diverse environmental conditions, *i.e.*, niches that lead to the divergence of microbial species. The phylogenetic niche conservatism theory predicts that two or more microbial species diverging from the same clade will have an overlap in their niches, implying that they are functionally redundant in some of their metabolic processes. The endogenous genetic factors that constrain the adaptation of individuals and, thus, populations to changing environmental conditions constitute the core process of phylogenetic niche conservatism. Furthermore, the degree of functional redundancy in a particular soil is proportional to the complexity of the considered function. We conclude with a conceptual model that identifies six patterns of functional redundancy in soil microbial communities, consistent with the phylogenetic niche conservatism theory.

Key Words: basic function, conceptual model, core gene, environmental factor, genetic factor, niche overlap, rare function, soil biodiversity

Citation: Jia Y, Whalen J K. 2020. A new perspective on functional redundancy and phylogenetic niche conservatism in soil microbial communities. *Pedosphere*. **30**(1): 18–24.

INTRODUCTION

Soil represents a vast reservoir of diverse microbial life, containing between 2 000 and 8.3 million bacterial species per gram soil (Gans *et al*., 2005; Schloss and Handelsman, 2006). Soil microbiota are responsible for degradation, transformation, and biosynthesis reactions in biogeochemical cycles as well as the detoxification of natural and human-made pollutants. The loss of a few species does not generally affect the soil functions because the same functions can be performed by multiple species (Jurburg and Salles, 2015; Grządziel, 2017). This functional redundancy could explain why the number of microbial species (alpha-diversity) is not related to soil functions (Wittebolle *et al*., 2008; Louca *et al*., 2016; Sheng *et al*., 2016).

The deliberate elimination or suppression of mi-

crobial groups has little effect on soil processes such as decomposition of organic matter (Griffiths *et al*., 2001a, b), indicating that multiple soil microbiota carry out the same functions (Grządziel, 2017; Nannipieri *et al*., 2017). Functional redundancy is common for some microbially-mediated processes. We can define functions as "basic" or "rare", depending on how frequently they occur in the soil environment. All soil microbiota have the metabolic capacity to perform "basic" soil functions such as respiration, nitrogen and phosphorus cycling, and organic matter decomposition (Wittebolle *et al.*, 2008; Nelson *et al.*, 2016; Grządziel, 2017). However, "rare" functions like methanogenesis or the mineralization of recalcitrant organic pollutants are mediated by specific groups, and there is far less redundancy in "rare" than "basic" functions (Girvan *et al*., 2005). Soil microbiota with the same functional

*[∗]*Corresponding author. E-mail: joann.whalen@mcgill.ca.

characteristics should have shared ancestry and have experienced similar adaptations to environmental conditions. This is consistent with the observations of Darwin (1859), which indicate that closely-related species share functional characteristics.

Suenaga (2015) used a metagenomics approach to compare the essential genes that encode enzymes for dissimilatory sulfate reduction and denitrification as well as the extradiol dioxygenases, which cleave the aromatic ring of catechol compounds during the aerobic microbial degradation of natural and xenobiotic aromatic compounds. The screening of thousands of clones revealed a high degree of similarity in the gene clusters of diverse prokaryotes, with some novel genes in the flanking regions. Mutations in the genes encoding such key enzymes would provide an adaptive phenotype optimized for a specific niche (Chattopadhyay *et al*., 2013). This is particularly evident in the extradiol dioxygenases, because the genes from a common ancestor that produced more thermostable extradiol dioxygenases have diverged through the nucleotide mutations, with modern lineages being less thermostable but possessing greater catabolic activity (Suenaga, 2015). Furthermore, Tringe *et al*. (2005) found that functional profiles of soil metagenomes were highly correlated with soil properties, suggesting that specific functions were selected by the environment (Louca *et al*., 2018).

Global warming and other anthropogenic factors are causing rapid environmental changes in all ecosystems, including the soil ecosystem. As microbialmediated processes are crucial to sustain soil functions, it is essential that we understand the root causes and consequences of functional redundancy of soil microbiota. Therefore, the objective of this perspective paper is to determine if the phylogenetic niche conservatism theory (Harvey and Pagel, 1991) could explain the functional redundancy of soil microbiota. The genetic and environmental factors leading to functional redundancy are presented, and the evidence for phylogenetic niche conservatism is shown in a conceptual model that describes the expected patterns of functional redundancy during the assembly of soil microbial communities.

PHYLOGENETIC NICHE CONSERVATISM AND FUNCTIONAL REDUNDANCY IN SOIL MICRO-BIAL COMMUNITIES

A fundamental concept in the phylogenetic niche conservatism theory is the definition of a niche, *i.e.*, the set of biotic and abiotic conditions in which a species

persists and maintains a stable population (Hutchinson, 1957). Two types of niches are distinguished: the fundamental niche and the realized niche (Hutchinson, 1957). The fundamental niche describes the abiotic conditions in which a species can persist, whereas the realized niche describes the conditions in which a species persists when other species (*e.g.*, competitors and predators) are present. As the fundamental niche can be conserved during evolutionary timespans, the phylogenetic niche conservatism theory predicts that modern lineages will have similar genetic, physiological, and ecological characteristics to their ancestors (Harvey and Pagel, 1991; Holt and Gaines, 1992; Wiens and Graham, 2005). Another aspect of the phylogenetic niche conservatism theory is that underlying biological processes constrain niche divergence between closely related species and, therefore, prevent populations from expanding into new niches (Losos, 2008).

The phylogenetic niche conservatism theory is consistent with the observed pattern of high niche similarity in related species, and it also explains the evolutionary mechanisms leading to that similarity (Losos, 2008; Wiens, 2008). As illustrated in Fig. 1, there are endogenous genetic, physiological, and ecological constraints that limit the ability of individuals and, thus, populations to rapidly adapt to changing environmental conditions (Pyron *et al*., 2015). Selective forces resulting from environmental variations experienced by a population tend to force the population to maintain their niche through time (Pyron *et al*., 2015). Three patterns are expected to emerge: niches are conserved (more similar than expected), constrained (divergent within a limited subset of available niches), or divergent (less similar than expected), on the basis of the degree of phylogenetic relatedness between species (Fig. 1).

Functional redundancy of soil microbiota is very likely a result of the first pattern, where niches are conserved and the species overlap due to a high degree of similarity at the genetic, physiological, and ecological levels (Fig. 1). Species that co-occur due to niche constraint (Fig. 1) are probably going to be functionally redundant as well. Although Fig. 1 illustrates niche overlap for two species, we expect multiple species to co-occur in soil niches as there are thousands to millions of species in soil microbial communities and an incalculable number of niches in the heterogeneous soil matrix. We also realize that niches are dynamic, not static, and the boundaries of a particular niche fluctuate with changing abiotic and biotic conditions, which allows population expansions and contractions in the niche (*i.e.*, population dynamics).

Fig. 1 Ecological and evolutionary consequences of phylogenetic niche conservatism for microbial communities. Endogenous factors constrain the phenotypes present, whereas exogenous factors represent selective pressures from ecological and environmental conditions that contribute to evolutionary processes. This leads to the three patterns of phylogenetic niche conservatism between species: niche conservatism, niche constraint, and niche divergence, where each species is represented as a circle and the niche is the dotted circle (adapted from Pyron *et al*. (2015)).

COMMON ANCESTORS WITH CORE METABO-LIC PROCESSES

The present-day soil microbial community is the outcome of the genetic and environmental adaptations of prokaryotes that descended from common ancestors on Earth. Their "basic" functions are the core metabolic processes that allow soil microbiota to alter the chemical speciation of virtually all elements on Earth (Falkowski *et al*., 2008). These metabolic processes involve the biogeochemical cycles of six major elements (hydrogen (H), carbon (C), nitrogen (N), oxygen (O), sulfur (S) , and phosphorus (P) that represent the major building blocks for all biological macromolecules (Schlesinger, 1997). These biogeochemical cycles have evolved on a planetary scale to form a set of nested, abiotically-driven acid-base reactions and biologicallydriven redox reactions (Falkowski *et al*., 2008), leading to a linked system of elemental cycles (Kluyver and Donker, 1926).

Core metabolic processes have survived intact and allowed prokaryotes to persist for billions of years, even though the Earth has undergone extraordinary environmental changes since its formation, such as the Great Oxygenation Event, ice ages, and massive volcanic outgassings (Knoll, 2003). Therefore, genes encoding for the core metabolic processes are highly conserved. These genes created and coevolved with biogeochemical cycles and were passed from microbe to microbe primarily by horizontal gene transfer (Falkowski *et al*., 2008; Suenaga, 2015). This is consistent with molecular biology studies based on gene order, which indicate that early cellular evolution was probably communal, with promiscuous horizontal gene flow as the principal mode of evolution (Woese, 2002). Core metabolic processes are controlled by gene clusters and families that are similar in thousands of organisms (Suenaga, 2015). In prokaryotes, gene homology is assured by horizontal gene transfer, *i.e.*, the movement of genetic material among organisms by transformation, transduction, and conjugation. This process often involves bacteriophages and plasmids (Varga *et al*., 2012, 2016) and differs from the vertical transmission of D-NA from parent to offspring (Robinson *et al*., 2013). In this manner, the genes responsible for the major extant catabolic and anabolic processes were distributed among organisms billions of years ago, before cellular differentiation and vertical gene transmission evolved as we know them today (Falkowski *et al*., 2008). Therefore, soil microbiota inevitably possesses genes for core metabolic processes within their DNA, which account for the functional redundancy in soil biogeochemical processes in modern-day microbial communities. Horizontal gene transfer is a historical evolutionary process that explains why diverse soil microbiota can perform many of the same functions. For instance, diverse groups of bacteria and archaea are able to fix atmospheric diatomic nitrogen (N_2) because they possess nitrogenase enzymes (Kechris *et al*., 2006). All ammonia-oxidizing bacteria and archaea produce ammonia monooxygenase, the key enzyme that oxidizes ammonia to hydroxylamine (Könneke *et al.*, 2005; Hallam *et al*., 2006). Furthermore, the sulfate-reducing Deltaproteobacteria, gram-positive bacteria, and Archaea share the same dissimilatory sulfite reductases that are responsible for sulfate respiration (Klein *et al*., 2001). As no single clade is responsible for these specific functions, prokaryotic phylogeny is weakly associated with N_2 fixation, ammonia oxidation, and sulfite reduction (Louca *et al*., 2018). Considering that these and other metabolic processes are not monophyletic (Aguilar *et al*., 2004; Martiny *et al*., 2013, 2015), it is

possible to find soil microbial communities with different species compositions that have a similar capacity to perform the same function (Fernández *et al.*, 1999; Turnbaugh *et al*., 2009; Burke *et al*., 2011). In fact, most Bacteria and Archaea lineages participate in biogeochemical cycling, although specialized reactions like photosynthesis and methanogenesis are limited to soil microbiota that possess the cellular machinery for these "rare" functions (Woese, 1987).

PHYLOGENETIC NICHE CONSERVATISM IS CO-NSISTENT WITH FUNCTIONAL REDUNDANCY DURING COMMUNITY ASSEMBLY

Microbial communities are expected to follow assembly rules, such that ecological processes selecting for or against species from the regional species pool will determine local community composition. These selective processes have been equated conceptually to hierarchical "filters" that act at increasingly finer scales to impose rules on the assemblage of communities (Keddy, 1992). Phylogenetic niche conservatism dominates the assembly process in the current soil microbial community, although horizontal gene transfer was more important in the early cellular evolution on the Earth (Woese, 2002). Global horizontal gene transfer is likely to have been relevant only to primitive genomes (Kurland *et al*., 2003). In modern organisms, both the range and frequency of horizontal gene transfer are constrained, most often by selective barriers (Kurland *et al*., 2003). The lineages (from ancestors to descendants) are the essence of genome evolution for contemporary organisms, therefore, horizontal gene transfer has a minimal impact on genome phylogeny in the current time period (Kurland *et al*., 2003). An analysis of genotypic and phenotypic data linked to 89 functions of Bacteria and Archaea in soil and other environmental matrices showed that 93% of the functions were nonrandomly distributed, which suggests that vertical inheritance was responsible for their functions (Martiny *et al*., 2015). Studies on the phylogenetic structure of communities of different major taxa and trophic levels, across different spatial and phylogenetic scales, revealed that 23 out of 39 studies (59%) found evidence of phylogenetic conservatism in contemporary communities (Vamosi *et al*., 2009).

Furthermore, at a phylogenetic level, microbial functions are conserved hierarchically, probably due to the complexity of the biochemical processes and the degree to which the functions are needed for survival (Martiny *et al*., 2015). For example, functions such as pH tolerance and salinity preference are deeply conserved

and shared among specialized taxa within deep clades (Philippot *et al*., 2010; Lennon *et al*., 2012; Placella *et al*., 2012). In contrast, the ability to use simple carbon substrates or absorb inorganic phosphorus are shallowly conserved, and generalist taxa from many clades are able to perform these functions (Martiny *et al*., 2015). The functional redundancy in "basic" functions of soil microbiota is therefore associated with inherited traits that respond to environmental factors, according to phylogenetic niche conservatism.

EXPECTED PATTERNS OF FUNCTIONAL RE-DUNDANCY DURING MICROBIAL COMMUNITY ASSEMBLY, BASED ON PHYLOGENETIC NICHE CONSERVATISM THEORY

The phylogenetic niche conservatism theory may explain the patterns of functional redundancy observed during microbial community assembly, as illustrated in Fig. 2. This conceptual model is suitable because it accounts for: i) evolutionary processes at the planetary scale, which produced a set of highly conserved core genes for redox reactions and biogeochemical cycles in all life-forms; ii) the genetic basis for the ongoing adaptation of microbiota within specific environments, *i.e.*, endogenous mutations; iii) the genetic and environmental factors that permit multiple species to co-occur within a niche or to diverge in neighboring niches, based on phylogenetic niche conservatism; and iv) the reasons for functional redundancy in "basic" functions and uniqueness of "rare" functions.

Fig. 2 Six patterns of functional redundancy for soil microbiota occupying the same niche. Two species are considered: microbe A and microbe B. The function is the biochemical process of transforming a substrate (State X) to a product (State Y). The reaction can proceed directly from State X to State Y (*i.e.*, microbe A1) or from State X to State Z before reaching State Y (*i.e.*, microbe A2). Dormant species are given a designation of zero (*i.e.*, microbes A_0 and B_0).

According to the phylogenetic niche conservatism theory, phylogenetically related microorganisms may be functional at the same time, which leads them to share the same functions when their niche overlaps $(A_1B_1$ pattern) until the environmental conditions change and their functions cease (Fig. 2). This is likely to occur when microbe A and microbe B are performing "basic" functions that both can do equally well. If microbe A is more efficient at acquiring resources or performing the function, it may be dominant and outcompete microbe B, resulting in the A_1B_0 pattern (Fig. 2). For their long-term survival in the overlap niche, microbe B has to have a deep phylogenetic trait such as the ability to perform the function when environmental conditions are no longer suitable for microbe A (*e.g.*, at the limits of A's tolerance for a particular stress, such as pH, salinity, oxygen, temperature, among others).

There are six possible patterns of functional redundancy for microbe A and microbe B (Fig. 2). These patterns reflect the ability of microbes to transform substrates, such as glucose (State X), to the reaction product, carbon dioxide (State Y). This is an abstract representation that should apply to any microbiallymediated reaction. Although a substrate can be transferred directly from State X to State Y, we acknowledge that other reactions may take place under certain environmental conditions, and produce an intermediate product such as pyruvic acid, State Z, that will eventually be converted to carbon dioxide, State Y (Fig. 2). Thus, there is more than one way to implement the same function. As soil microbiota are ubiquitous, but often are present in dormant or resting states, awaiting an opportunity to "bloom" (Jurburg and Salles, 2015), this model allows for co-occurrence of active and dormant species within the same niche. For instance, the A_1A_2 and A_1A_0 patterns suggest that microbe A is active and transforms State X to State Y directly (A_1) or through another pathway (A_2) transfers the substrate from State X to State Z, then State Y), whereas microbe B remains dormant. This is obviously a simplistic view of soil microbiota functions, but it explains how functional redundancy occurs within the context of the phylogenetic niche conservatism theory.

PERSPECTIVES AND FUTURE DIRECTIONS

Functional redundancy of soil microbiota has developed through evolution and reflects local adaptation to environmental conditions. It seems likely that soil microbial community assembly follows the principles of phylogenetic niche conservatism, but this remains to be proven experimentally. Soil metagenomics studies allow for rapid characterization of thousands of soil microbiomes and should prove helpful for establishing the phylogenetic supertree representing the species assembled in a particular community. Evaluating the functional genes of the soil microbiota should provide more insights to understand functional redundancy as well as the occurrence of "rare" functions within soil microbial communities.

Functional redundancy in soil microbiota is widely accepted, but generally not explained. A deeper understanding of soil biodiversity requires a theoretical basis to resolve many ecological and evolutionary issues, such as the co-existence of species in the heterogeneous soil matrix, the relationship between community structure and function, among others. Many theories could be considered as potential explanations of this phenomenon, such as neutral theory, trade-off, gene drift, high dimensional niches, fluctuating selection, dormant cells, or patchy environmental conditions, but none of these are fully accepted by ecologists. We propose the phylogenetic niche conservatism theory simply because it provides a better explanation of our understanding of the issue of functional redundancy. It remains to be confirmed whether this is the best theory for explaining functional redundancy in soil microbial communities.

ACKNOWLEDGEMENTS

This work was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC) through Discovery Grant (No. RGPIN-2017- 05391). We thank the Soil Ecology Research Group members for helpful discussion and suggestions.

REFERENCES

- Aguilar D, Aviles F X, Querol E, Sternberg M J E. 2004. Analysis of phenetic trees based on metabolic capabilites across the three domains of life. *J Mol Biol*. **340**: 491–512.
- Burke C, Steinberg P, Rusch D, Kjelleberg S, Thomas T. 2011. Bacterial community assembly based on functional genes rather than species. *Proc Natl Acad Sci USA*. **108**: 14288– 14293.
- Chattopadhyay S, Taub F, Paul S, Weissman S J, Sokurenko E V. 2013. Microbial variome database: Point mutations, adaptive or not, in bacterial core genomes. *Mol Biol Evol*. **30**: 1465–1470.
- Darwin C. 1859. On the Origin of Species. Harvard University Press, Cambridge.
- Falkowski P G, Fenchel T, Delong E F. 2008. The microbial engines that drive Earth's biogeochemical cycles. *Science*. **320**: 1034–1039.
- Fernández A, Huang S Y, Seston S, Xing J, Hickey R, Criddle C, Tiedje J. 1999. How stable is stable? Function *versus* community composition. *Appl Environ Microbiol*. **65**: 3697– 3704.
- Gans J, Wolinsky M, Dunbar J. 2005. Computational improvements reveal great bacterial diversity and high metal toxicity in soil. *Science*. **309**: 1387–1390.
- Girvan M S, Campbell C D, Killham K, Prosser J I, Glover L A. 2005. Bacterial diversity promotes community stability and functional resilience after perturbation. *Environ Microbiol*. **7**: 301–313.
- Griffiths B S, Bonkowski M, Roy J, Ritz K. 2001a. Functional stability, substrate utilisation and biological indicators of soils following environmental impacts. *Appl Soil Ecol*. **16**: 49–61.
- Griffiths B S, Ritz K, Wheatley R, Kuan H L, Boag B, Christensen S, Ekelund F, Sørensen S J, Muller S, Bloem J. 2001b. An examination of the biodiversity—ecosystem function relationship in arable soil microbial communities. *Soil Biol Biochem*. **33**: 1713–1722.
- Grządziel J. 2017. Functional redundancy of soil microbiotadoes more always mean better? *Pol J Soil Sci*. **50**: 75–81.
- Hallam S J, Mincer T J, Schleper C, Preston C M, Roberts K, Richardson P M, DeLong E F. 2006. Pathways of carbon assimilation and ammonia oxidation suggested by environmental genomic analyses of marine *Crenarchaeota*. *PLoS Biol*. **4**: e95.
- Harvey P H, Pagel M D. 1991. The Comparative Method in Evolutionary Biology. Oxford University Press, Oxford.
- Holt R D, Gaines M S. 1992. Analysis of adaptation in heterogeneous landscapes: Implications for the evolution of fundamental niches. *Evol Ecol*. **6**: 433–447.
- Hutchinson G E. 1957. Concluding remarks. *Cold Spring Harb Symp Quant Biol*. **22**: 415–427.
- Jurburg S D, Salles J F. 2015. Functional redundancy and ecosystem function—the soil microbiota as a case study. *In* Yueh-Hsin L, Blanco J A, Shovonlal R (eds.) Ecosystems—linking Structure and Function. InTech Open Science, Rijeka. pp. 29–42.
- Kechris K J, Lin J C, Bickel P J, Glazer A N. 2006. Quantitative exploration of the occurrence of lateral gene transfer by using nitrogen fixation genes as a case study. *Proc Natl Acad Sci USA*. **103**: 9584–9589.
- Keddy P A. 1992. Assembly and response rules: Two goals for predictive community ecology. *J Veg Sci*. **3**: 157–164.
- Klein M, Friedrich M, Roger A J, Hugenholtz P, Fishbain S, Abicht H, Blackall L L, Stahl D A, Wagner M. 2001. Multiple lateral transfers of dissimilatory sulfite reductase genes between major lineages of sulfate-reducing prokaryotes. *J Bacteriol*. **183**: 6028–6035.
- Kluyver A J, Donker H J K. 1926. Die einheit in der biochemie (The Unit in Biochemistry). *Chem Zelle Gewebe* (in German). **13**: 134–190.
- Knoll A H. 2003. Life on a Young Planet: The First Three Billion Years of Evolution on Earth. Princeton University Press, Princeton.
- Könneke M, Bernhard A E, de la Torre J R, Walker C B, Waterbury J B, Stahl D A. 2005. Isolation of an autotrophic ammonia-oxidizing marine archaeon. *Nature*. **437**: 543–546.
- Kurland C G, Canback B, Berg O G. 2003. Horizontal gene transfer: A critical view. *Proc Natl Acad Sci USA*. **100**: 9658–9662.
- Lennon J T, Aanderud Z T, Lehmkuhl B K, Schoolmaster Jr D R. 2012. Mapping the niche space of soil microorganisms using taxonomy and traits. *Ecology*. **93**: 1867–1879.
- Losos J B. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecol Lett*. **11**: 995– 1003.
- Louca S, Jacques S M S, Pires A P F, Leal J S, Srivastava D S, Parfrey L W, Farjalla V F, Doebeli M. 2016. High taxonomic variability despite stable functional structure across

microbial communities. *Nat Ecol Evol*. **1**: 0015.

- Louca S, Polz M F, Mazel F, Albright M B N, Huber J A, O'Connor M I, Ackermann M, Hahn A S, Srivastava D S, Crowe S A, Doebeli M, Parfrey L W. 2018. Function and functional redundancy in microbial systems. *Nat Ecol Evol*. **2**: 936–943.
- Martiny A C, Treseder K, Pusch G. 2013. Phylogenetic conservatism of functional traits in microorganisms. *ISME J*. **7**: 830–838.
- Martiny J B H, Jones S E, Lennon J T, Martiny A C. 2015. Microbiomes in light of traits: A phylogenetic perspective. *Science*. **350**: aac9323.
- Nannipieri P, Ascher J, Ceccherini M T, Landi L, Pietramellara G, Renella G. 2017. Microbial diversity and soil functions. *Eur J Soil Sci*. **68**: 12–26.
- Nelson M B, Martiny A C, Martiny J B H. 2016. Global biogeography of microbial nitrogen-cycling traits in soil. *Proc Natl Acad Sci USA*. **113**: 8033–8040.
- Philippot L, Andersson S G E, Battin T J, Prosser J I, Schimel J P, Whitman W B, Hallin S. 2010. The ecological coherence of high bacterial taxonomic ranks. *Nat Rev Microbiol*. **8**: 523– 529.
- Placella S A, Brodie E L, Firestone M K. 2012. Rainfall-induced carbon dioxide pulses result from sequential resuscitation of phylogenetically clustered microbial groups. *Proc Natl Acad Sci USA*. **109**: 10931–10936.
- Pyron R A, Costa G C, Patten M A, Burbrink F T. 2015. Phylogenetic niche conservatism and the evolutionary basis of ecological speciation. *Biol Rev*. **90**: 1248–1262.
- Robinson K M, Sieber K B, Hotopp J C D. 2013. A review of bacteria-animal lateral gene transfer may inform our understanding of diseases like cancer. *PLoS Genet*. **9**: e1003877.
- Schlesinger W H. 1997. Biogeochemistry: An Analysis of Global Change. 2nd Edn. Academic Press, New York.
- Schloss P D, Handelsman J. 2006. Toward a census of bacteria in soil. *PLoS Comput Biol*. **2**: e92.
- Sheng Y Z, Bibby K, Grettenberger C, Kaley B, Macalady J L, Wang G C, Burgos W D. 2016. Geochemical and temporal influences on the enrichment of acidophilic iron-oxidizing bacterial communities. *Appl Environ Microbiol*. **82**: 3611– 3621.
- Suenaga H. 2015. Targeted metagenomics unveils the molecular basis for adaptive evolution of enzymes to their environment. *Front Microbiol*. **6**: 1018.
- Tringe S G, von Mering C, Kobayashi A, Salamov A A, Chen K, Chang H W, Podar M, Short J M, Mathur E J, Detter J C, Bork P, Hugenholtz P, Rubin E M. 2005. Comparative metagenomics of microbial communities. *Science*. **308**: 554–557.
- Turnbaugh P J, Hamady M, Yatsunenko T, Cantarel B L, Duncan A, Ley R E, Sogin M L, Jones W J, Roe B A, Affourtit J P, Egholm M, Henrissat B, Heath A C, Knight R, Gordon J I. 2009. A core gut microbiome in obese and lean twins. *Nature*. **457**: 480–484.
- Vamosi S M, Heard S B, Vamosi J C, Webb C O. 2009. Emerging patterns in the comparative analysis of phylogenetic community structure. *Mol Ecol*. **18**: 572–592.
- Varga M, Kuntová L, Pantůček R, Mašlaňová I, Růžičková V, Doškař J. 2012. Efficient transfer of antibiotic resistance plasmids by transduction within methicillin-resistant *Staphylococcus aureus* USA300 clone. *FEMS Microbiol Lett*. **332**: 146– 152.
- Varga M, Pantůček R, Růžičková V, Doškař J. 2016. Molecular characterization of a new efficiently transducing bacteriophage identified in meticillin-resistant *Staphylococcus aureus*. *J Gen Virol*. **97**: 258–268.
- Wiens J J. 2008. Commentary on Losos (2008): Niche conservatism déjà vu. *Ecol Lett*. **11**: 1004–1005.
- Wiens J J, Graham C H. 2005. Niche conservatism: Integrating evolution, ecology, and conservation biology. *Annu Rev Ecol Evol Syst*. **36**: 519–539.
- Wittebolle L, Vervaeren H, Verstraete W, Boon N. 2008. Quanti-

fying community dynamics of nitrifiers in functionally stable reactors. *Appl Environ Microbiol*. **74**: 286–293.

- Woese C R. 1987. Bacterial evolution. *Microbiol Rev*. **51**: 221– 271.
- Woese C R. 2002. On the evolution of cells. *Proc Natl Acad Sci USA*. **99**: 8742–8747.